

**SITE-DIRECTED MUTAGENESIS OF SUPERFOLDER  
GREEN FLUORESCENT PROTEIN**

By

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## **ABSTRACT**

### **SITE-DIRECTED MUTAGENESIS OF SUPERFOLDER GREEN FLUORESCENT PROTEIN**

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Green Fluorescent Protein (GFP) is a protein made up of 238 amino acid residues that will emit green fluorescence when it is exposed to blue or UV light. It consists of eleven antiparallel beta-strands forming a beta-barrel, with an alpha-helix inside. It has been employed extensively as a reporter gene to visualize the protein-protein interactions (PPI) using GFP in bimolecular fluorescence complementation (BiFC) assays. Numerous researches have been carried out to develop GFP variants, including superfolder GFP (sfGFP), for improving the signal-to-noise (S/N) ratio in the BiFC assays, due to the intrinsic intramolecular interaction of the beta-strands, which causes irreversible assembly of sfGFP in the assays. This limits the use of sfGFP for monitoring dynamic PPI. The main objective of this study is to clone and express variants of sfGFP with mutation at 8th beta sheet and examine if these mutants sfGFP would remain fluorescent. Mutations in the 8th beta sheet of sfGFP, were separately introduced by polymerase chain reaction (PCR), generating DNA fragments of mutant C-terminal of sfGFP (sfGFP-C).

Subsequently, the N-terminal fragment of sfGFP, was separately fused to the mutant sfGFP-C fragments by overlap extension polymerase chain reaction (OE-PCR). The full-length mutant sfGFP fragments were cloned into the expression plasmid vector pBTOPO<sup>®</sup> and the reaction mixtures were used to transform competent *Escherichia coli* TOP10. Transformants were screened by blue light irradiation, where putative carrying recombinant plasmid with the sfGFP insert would emit green fluorescence. In summary, all five mutant sfGFP constructs were separately cloned into the pB-TOPO<sup>®</sup>. However, mutant sfGFP IV was inserted into the expression vector at the opposite orientation, and therefore, was non-functional. Mutant sfGFP exhibited fluorescence loss, while other mutants, produced lower fluorescence than that of the wild-type sfGFP.

## **ACKNOWLEDGEMENT**

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## DECLARATION

I hereby declare that the project report is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UTAR or other institutions.

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LEE SOCK IM

## APPROVAL SHEET

This project report entitled “**SITE-DIRECTED MUTAGENESIS OF SUPERFOLDER GREEN FLUORESCENT PROTEIN**” was prepared by LEE SOCK IM and submitted as partial fulfilment of the requirements for the degree of Bachelor of Science (Hons) Biotechnology at Universiti Tunku Abdul Rahman.

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**PERMISSION SHEET**

It is hereby certified that **LEE SOCK IM** (ID No: **12ADB01797**) has completed this final year project entitled **“SITE-DIRECTED MUTAGENESIS OF SUPERFOLDER GREEN FLUOROSCENT PROTEIN”** under the supervision of Assoc. Prof. Dr. Wong Hann Ling from the Department of Biological Science, Faculty of Science.

I hereby give permission to the University to upload the softcopy of my final year project in pdf format into the UTAR Institutional Repository, which may be made accessible to the UTAR community and public.

Yours truly,

\_\_\_\_\_  
(LEE SOCK IM)

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